

# Package ‘rmsb’

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**Title** Bayesian Regression Modeling Strategies

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## Description

A Bayesian companion to the 'rms' package, 'rmsb' provides Bayesian model fitting, post-fit estimation, and graphics. It implements Bayesian regression models whose fit objects can be processed by 'rms' functions such as 'contrast()', 'summary()', 'Predict()', 'nomogram()', and 'latex()'. The fitting function currently implemented in the package is 'blrm()' for Bayesian logistic binary and ordinal regression with optional clustering, censoring, and departures from the proportional odds assumption using the partial proportional odds model of Peterson and Harrell (1990) <<https://www.jstor.org/stable/2347760>>.

**License** GPL (>= 3)

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rmsb-package

*The 'rmsb' package.*

---

## Description

Regression Modeling Strategies Bayesian

The **rmsb** package is an appendage to the **rms** package that implements Bayesian regression models whose fit objects can be processed by **rms** functions such as `contrast`, `summary`, `Predict`, `nomogram`, and `latex`. The fitting function currently implemented in the package is `blrm` for Bayesian logistic binary and ordinal regression with optional clustering, censoring, and departures from the proportional odds assumption using the partial proportional odds model of Peterson and Harrell (1990).

## References

Stan Development Team (2020). RStan: the R interface to Stan. R package version 2.19.3. <https://mc-stan.org>

## See Also

- <https://hbiostat.org/R/rmsb/> for the package's main web page
- <https://hbiostat.org/R/examples/blrm/blrm.html> for a vignette with many examples of using the `blrm` function

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blrm

*Bayesian Binary and Ordinal Logistic Regression*

---

## Description

Uses `rstan` with pre-compiled Stan code, or `cmdstan` to get posterior draws of parameters from a binary logistic or proportional odds semiparametric ordinal logistic model. The Stan code internally using the qr decomposition on the design matrix so that highly collinear columns of the matrix do not hinder the posterior sampling. The parameters are transformed back to the original scale before returning results to R. Design matrix columns are centered before running Stan, so Stan diagnostic output will have the intercept terms shifted but the results of `blrm()` for intercepts are for the original uncentered data. The only prior distributions for regression betas are normal with mean zero. Priors are specified on contrasts so that they can be specified on a meaningful scale and so that more complex patterns can be imposed. Parameters that are not involved in any contrasts in `pcontrast` or `npcontrast` have non-informative priors. Contrasts are automatically converted to the QR space used in Stan code.

**Usage**

```

blrm(
  formula,
  ppo = NULL,
  cppo = NULL,
  data = environment(formula),
  subset,
  na.action = na.delete,
  priorsdppo = rep(100, pppo),
  iprior = 0,
  conc = 1/(0.8 + 0.35 * max(k, 3)),
  ascale = 1,
  psigma = 1,
  rsdmean = if (psigma == 1) 0 else 1,
  rsdsd = 1,
  normcppo = FALSE,
  pcontrast = NULL,
  npcontrast = NULL,
  backend = c("rstan", "cmdstan"),
  iter = 2000,
  warmup = iter/2,
  chains = 4,
  refresh = 0,
  progress = if (refresh > 0) "stan-progress.txt" else "",
  x = TRUE,
  y = TRUE,
  loo = n <= 1000,
  ppairs = NULL,
  method = c("both", "sampling", "optimizing"),
  inito = if (length(ppo)) 0 else "random",
  inits = inito,
  standata = FALSE,
  file = NULL,
  debug = FALSE,
  sampling.args = NULL,
  showopt = FALSE,
  ...
)

```

**Arguments**

formula	a R formula object that can use rms package enhancements such as the restricted interaction operator
ppo	formula specifying the model predictors for which proportional odds is not assumed
cppo	a function that if present causes a constrained partial PO model to be fit. The function specifies the values in the Gamma vector in Peterson and Harrell (1990)

equation (6). Sometimes to make posterior sampling better behaved, the function should be scaled and centered. This is done by wrapping `cppo` in a function that scales the `cppo` result before returning the vector value, when `normcppo` is `TRUE`. The default normalization is based on the mean and standard deviation of the function values over the distribution of observed `Y`. For getting predicted values and estimates post-`blrm()`, `cppo` must not reference any functions that are not available at such later times.

<code>data</code>	a data frame; defaults to using objects from the calling environment
<code>subset</code>	a logical vector or integer subscript vector specifying which subset of data should be used
<code>na.action</code>	default is <code>na.delete</code> to remove missings and report on them
<code>priorsdppo</code>	vector of prior standard deviations for non-proportional odds parameters. The last element is the only one for which the SD corresponds to the original data scale. This only applies to the unconstrained PPO model.
<code>iprior</code>	specifies whether to use a Dirichlet distribution for the cell probabilities, which induce a more complex prior distribution for the intercepts ( <code>iprior=0</code> , the default), non-informative priors ( <code>iprior=1</code> ) directly on the intercept parameters, or to directly use a t-distribution with 3 d.f. and scale parameter <code>ascale</code> ( <code>iprior=2</code> ).
<code>conc</code>	the Dirichlet distribution concentration parameter for the prior distribution of cell probabilities at covariate means. The default is the reciprocal of $0.8 + 0.35 \max(k, 3)$ where $k$ is the number of <code>Y</code> categories. The default is chosen to make the posterior mean of the intercepts more closely match the MLE. For optimizing, the concentration parameter is always 1.0 when <code>iprior=0</code> to obtain results very close to the MLE for providing the posterior mode.
<code>ascale</code>	scale parameter for the t-distribution for priors for the intercepts if <code>iprior=2</code> , defaulting to 1.0
<code>psigma</code>	defaults to 1 for a half-t distribution with 4 d.f., location parameter <code>rsdmean</code> and scale parameter <code>rsdsd</code> . Set <code>psigma=2</code> to use the exponential distribution.
<code>rsdmean</code>	the assumed mean of the prior distribution of the standard deviation of random effects. When <code>psigma=2</code> this is the mean of an exponential distribution and defaults to 1. When <code>psigma=1</code> this is the mean of the half-t distribution and defaults to zero.
<code>rsdsd</code>	applies only to <code>psigma=1</code> and is the scale parameter for the half t distribution for the SD of random effects, defaulting to 1.
<code>normcppo</code>	set to <code>TRUE</code> to modify the <code>cppo</code> function automatically centering and scaling the result
<code>pcontrast</code>	a list specifying contrasts that are to be given Gaussian prior distributions. The predictor combinations specified in <code>pcontrast</code> are run through <code>rms::gendata()</code> so that contrasts are specified in units of original variables, and unspecified variables are set to medians or modes as saved by <code>rms::datadist()</code> . Thanks to Stan, putting priors on combinations and transformations of model parameters has the same effect of putting different priors on the original parameters without figuring out how to do that. The syntax used here allows specification of differences, double differences (e.g., interactions or nonlinearity), triple differences (e.g., to put constraints on nonlinear interactions), etc. The requested predictor

combinations must be named so they may be referred to inside `contrast`. The syntax is `pcontrast=list(..., contrast=expression(...), mu=, sd=, weights=, expand=)`. `...` denotes one or more `list()`s with predictor combinations, and each `list()` must be named, e.g., `pcontrast=list(c1=list(sex='female'), c2=list(sex='male'))` to set up for a female - male contrast specified as `contrast=expression(c1 - c2)`. The `c1 - c2` subtraction will operate on the design matrices generated by the covariate settings in the `list()`s. For `weights`, `expand` see `rms::Xcontrast()` and `rms::contrast.rms()`. `mu` is a vector of prior means associated with the rows of the stacked contrasts, and `sd` is a corresponding vector of Gaussian prior SDs. When `mu` is not given it defaults to 0.0, and `sd` defaults to 100.0. Values of `mu` and/or `sd` are repeated to the number of contrasts if they are of length 1. Full examples are given [here](#).

<code>npcontrast</code>	like <code>pcontrast</code> but applies to the non-proportional odds submodel in <code>ppo</code> . Priors for the amount of departure from proportional odds are isolated from the priors of the "main effects" in formula. The mean and standard deviation for the non-PO contrasts are on the scale of $Z \cdot \tau$ before <code>cppo</code> is applied. If <code>cppo</code> picks off a single condition, i.e., death is the highest level of <code>Y</code> and you want a special effect of treatment on death, then <code>cppo</code> will be something like <code>function(y) y == 4</code> and the contrast prior will be on the scale of the additional treatment effect for death. If <code>cppo</code> is more of a continuous function you will have to take into account the values of that function when figuring the prior mean and SD. For example, if <code>y</code> ranges from 10-90 and <code>cppo</code> is <code>sqrt(y)</code> , and you want to specify a prior on the log odds ratio for <code>y=10</code> vs. <code>y=90</code> you'll need to divide the prior standard deviation in <code>npcontrast</code> by <code>sqrt(90) - sqrt(10)</code> .
<code>backend</code>	set to <code>cmdstan</code> to use <code>cmdstan</code> through the R <code>cmdstanr</code> package instead of the default <code>rstan</code> . You can also specify this with a global option <code>rmsb.backend</code> .
<code>iter</code>	number of posterior samples per chain for <code>rstan::sampling()</code> to run, counting warmups
<code>warmup</code>	number of warmup iterations to discard. Default is <code>iter/2</code> .
<code>chains</code>	number of separate chains to run
<code>refresh</code>	see <code>rstan::sampling()</code> and <code>cmdstanr::sample()</code> . The default is 0, indicating that no progress notes are output. If <code>refresh &gt; 0</code> and progress is not <code>' '</code> , progress output will be appended to file <code>progress</code> . The default file name is <code>'stan-progress.txt'</code> .
<code>progress</code>	see <code>refresh</code> . Defaults to <code>' '</code> if <code>refresh = 0</code> . Note: If running interactively but not under RStudio, <code>rstan</code> will open a browser window for monitoring progress.
<code>x</code>	set to <code>FALSE</code> to not store the design matrix in the fit. <code>x=TRUE</code> is needed if running <code>blrmStats</code> for example.
<code>y</code>	set to <code>FALSE</code> to not store the response variable in the fit
<code>loo</code>	set to <code>FALSE</code> to not run <code>loo</code> and store its result as object <code>loo</code> in the returned object. <code>loo</code> defaults to <code>FALSE</code> if the sample size is greater than 1000, as <code>loo</code> requires the per-observation likelihood components, which creates a matrix <code>N</code> times the number of posterior draws.
<code>ppairs</code>	set to a file name to run <code>rstan</code> pairs or, if <code>backend='cmdstan'</code> <code>bayesplot::mcmc_pairs</code> and store the resulting png plot there. Set to <code>TRUE</code> instead to directly plot these diagnostics. The default is not to run pair plots.

method	set to 'optimizing' to run the Stan optimizer and not do posterior sampling, 'both' (the default) to run both the optimizer and posterior sampling, or 'sampling' to run only the posterior sampling and not compute posterior modes. Running optimizing is a way to obtain maximum likelihood estimates and allows one to quickly study the effect of changing the prior distributions. When method='optimizing' is used the result returned is not a standard <code>blrm()</code> object but is instead the parameter estimates, -2 log likelihood, and optionally the Hessian matrix (if you specify <code>hessian=TRUE</code> in ...; not available with <code>cmdstan</code> ). When method='both' is used, <code>rstan::sampling()</code> and <code>rstan::optimizing()</code> are both run, and parameter estimates (posterior modes) from optimizing are stored in a matrix <code>param</code> in the fit object, which also contains the posterior means and medians, and other results from optimizing are stored in object <code>opt</code> in the <code>blrm()</code> fit object. When random effects are present, method is automatically set to 'sampling' as maximum likelihood estimates without marginalizing over the random effects do not make sense. When you specify method='optimizing' specify <code>iprior=</code> to get regular MLEs in which no prior is put on the intercepts.
inito	initial value for optimization. The default is the <code>rstan</code> default 'random'. Frequently specifying <code>init=0</code> will benefit when the number of distinct Y categories grows or when using <code>ppo</code> hence 0 is the default for that.
inits	initial value for sampling, defaults to <code>inito</code>
standata	set to <code>TRUE</code> to return the Stan data list and not run the model
file	a file name for a <code>saveRDS</code> -created file containing or to contain the saved fit object. If <code>file</code> is specified and the file does not exist, it will be created right before the fit object is returned, less the large <code>rstan</code> object. If the file already exists, its stored md5 hash string <code>datahash</code> fit object component is retrieved and compared to that of the current <code>rstan</code> inputs. If the data to be sent to <code>rstan</code> , the priors, and all sampling and optimization options and stan code are identical, the previously stored fit object is immediately returned and no new calculations are done.
debug	set to <code>TRUE</code> to output timing and progress information to <code>/tmp/debug.txt</code>
sampling.args	a list containing parameters to pass to <code>rstan::sampling()</code> or to the <code>rcmdstan</code> sample function, other than these arguments: <code>iter</code> , <code>warmup</code> , <code>chains</code> , <code>refresh</code> , <code>init</code> which are already arguments to <code>blrm</code> . A good use of this is <code>sampling.args=list(seed=3)</code> to get reproducible sampling.
showopt	set to <code>TRUE</code> to show Stan optimizer output
...	passed to <code>rstan::optimizing()</code> or the <code>rcmdstan</code> optimizing function. <code>sampling.args</code> is usually used instead.

## Details

The partial proportional odds model of Peterson and Harrell (1990) is implemented, and is invoked when the user specifies a second model formula as the `ppo` argument. This formula has no left-hand-side variable, and has right-side variables that are a subset of those in formula specifying for which predictors the proportional odds assumption is relaxed.

The Peterson and Harrell (1990) constrained partial proportional odds is also implemented, and is usually preferred to the above unconstrained PPO model as it adds a vector of coefficients instead of a matrix of coefficients. In the constrained PPO model the user provides a function `cppo` that

computes a score for all observed values of the dependent variable. For example with a discrete ordinal outcome `cppo` may return a value of 1.0 for a specific value of `Y` and zero otherwise. That will result in a departure from the proportional odds assumption for just that one level of `Y`. The value returned by `cppo` at the lowest `Y` value is never used in any case.

`blrm()` also handles single-level hierarchical random effects models for the case when there are repeated measurements per subject which are reflected as random intercepts, and a different experimental model that allows for AR(1) serial correlation within subject. For both setups, a `cluster` term in the model signals the existence of subject-specific random effects.

When using the `cmdstan` backend, `cmdstanr` will need to compile the Stan code once per computer, only recompiling the code when the Stan source code changes. By default the compiled code is stored in directory `.rmsb` under your home directory. Specify `options(rmsbdir=)` to specify a different location. You should specify `rmsbdir` to be in a project-specific location if you want to archive code for old projects.

If you want to run MCMC sampling even when no inputs or Stan code have changed, i.e., to use a different random number seed for the sampling process when you did not specify `sampling.args(seed=...)`, remove the file before running `blrm`.

Set `options(rmsbmsg=FALSE)` to suppress certain information messages.

See [here](#) and [here](#) for multiple examples with results.

## Value

an `rms` fit object of class `blrm`, `rmsb`, `rms` that also contains `rstan` or `cmdstanr` results under the name `rstan`. In the `rstan` results, which are also used to produce diagnostics, the intercepts are shifted because of the centering of columns of the design matrix done by `blrm()`. With `method='optimizing'` a class-less list is returned with these elements: `coefficients` (MLEs), `beta` (non-intercept parameters on the QR decomposition scale), `deviance` ( $-2 \log$  likelihood), `return_code` (see `rstan::optimizing()`), and, if you specified `hessian=TRUE` to `blrm()`, the Hessian matrix. To learn about the scaling of orthogonalized QR design matrix columns, look at the `xqrstd` object in the returned object. This is the vector of SDs for all the columns of the transformed matrix. The returned element `sampling_time` is the elapsed time for running posterior samplers, in seconds. This will be just a little more than the time for running one CPU core for one chain.

## Author(s)

Frank Harrell and Ben Goodrich

## See Also

`print.blrm()`, `blrmStats()`, `stanDx()`, `stanGet()`, `coef.rmsb()`, `vcov.rmsb()`, `print.rmsb()`, `coef.rmsb()`

## Examples

```
## Not run:
getHdata(titanic3)
dd <- datadist(titanic3); options(datadist='dd')
f <- blrm(survived ~ (rcs(age, 5) + sex + pclass)^2, data=titanic3)
```



```

f                # model summary using print.blrm
coef(f)          # compute posterior mean parameter values
coef(f, 'median') # compute posterior median values
stanDx(f)        # print basic Stan diagnostics
s <- stanGet(f)  # extract rstan object from fit
plot(s, pars=f$betas) # Stan posteriors for beta parameters
stanDxplot(s)    # Stan diagnostic plots by chain
blrmStats(f)     # more details about predictive accuracy measures
ggplot(Predict(...)) # standard rms output
summary(f, ...)  # invokes summary.rms
contrast(f, ...) # contrast.rms computes HPD intervals
plot(nomogram(f, ...)) # plot nomogram using posterior mean parameters

# Fit a random effects model to handle multiple observations per
# subject ID using cmdstan
# options(rmsb.backend='cmdstan')
f <- blrm(outcome ~ rcs(age, 5) + sex + cluster(id), data=mydata)

## End(Not run)

```

---

blrmStats

---

*Compute Indexes of Predictive Accuracy and Their Uncertainties*


---

## Description

For a binary or ordinal logistic regression fit from `blrm()`, computes several indexes of predictive accuracy along with highest posterior density intervals for them. Optionally plots their posterior densities. When there are more than two levels of the outcome variable, computes Somers' Dxy and c-index on a random sample of 10,000 observations.

## Usage

```
blrmStats(fit, ns = 400, prob = 0.95, pl = FALSE, dist = c("density", "hist"))
```

## Arguments

fit	an object produced by <code>blrm()</code>
ns	number of posterior draws to use in the calculations (default is 400)
prob	HPD interval probability (default is 0.95)
pl	set to TRUE to plot the posterior densities using base graphics
dist	if pl is TRUE specifies whether to plot the density estimate (the default) or a histogram

**Value**

list of class `blrmStats` whose most important element is `Stats`. The indexes computed are defined below, with `gp`, `B`, `EV`, and `vp` computed using the intercept corresponding to the median value of `Y`. See <https://fharrell.com/post/addvalue> for more information.

**"Dxy"** Somers' Dxy rank correlation between predicted and observed. The concordance probability (c-index; AUROC in the binary `Y` case) may be obtained from the relationship  $Dxy=2(c-0.5)$ .

**"g"** Gini's mean difference: the average absolute difference over all pairs of linear predictor values

**"gp"** Gini's mean difference on the predicted probability scale

**"B"** Brier score

**"EV"** explained variation

**"v"** variance of linear predictor

**"vp"** variable of estimated probabilities

**Author(s)**

Frank Harrell

**See Also**

[Hmisc::rcorr.cens\(\)](#)

**Examples**

```
## Not run:
f <- blrm(...)
blrmStats(f, pl=TRUE) # print and plot

## End(Not run)
```

---

cluster

*cluster*

---

**Description**

Cluster Function for Random Effects

**Usage**

```
cluster(x)
```

**Arguments**

`x` a vector representing a categorical vector

**Details**

Used by blrm to signal a categorical variable to generate random effects.

**Value**

x unchanged

**Author(s)**

Frank Harrell

---

 coef.rmsb

*Extract Bayesian Summary of Coefficients*


---

**Description**

Computes either the posterior mean (default), posterior median, or posterior mode of the parameters in an rms Bayesian regression model

**Usage**

```
## S3 method for class 'rmsb'
coef(object, stat = c("mean", "median", "mode"), ...)
```

**Arguments**

object	an object created by an rms package Bayesian fitting function
stat	name of measure of posterior distribution central tendency to compute
...	ignored

**Value**

a vector of intercepts and regression coefficients

**Author(s)**

Frank Harrell

**Examples**

```
## Not run:
f <- blrm(...)
coef(f, stat='mode')

## End(Not run)
```

---

<code>compareBmods</code>	<i>Compare Bayesian Model Fits</i>
---------------------------	------------------------------------

---

**Description**

Uses `loo::loo_model_weights()` to compare a series of models such as those created with `blrm()`

**Usage**

```
compareBmods(..., method = "stacking", r_eff_list = NULL)
```

**Arguments**

<code>...</code>	a series of model fits
<code>method</code>	see <code>loo::loo_model_weights()</code>
<code>r_eff_list</code>	see <code>loo::loo_model_weights()</code>

**Value**

a `loo::loo_model_weights()` object

**Author(s)**

Frank Harrell

---

<code>distSym</code>	<i>Distribution Symmetry Measure</i>
----------------------	--------------------------------------

---

**Description**

From a sample from a distribution computes a symmetry measure. By default it is the gap between the mean and the 0.95 quantile divided by the gap between the 0.05 quantile and the mean.

**Usage**

```
distSym(x, prob = 0.9, na.rm = FALSE)
```

**Arguments**

<code>x</code>	a numeric vector representing a sample from a continuous distribution
<code>prob</code>	quantile interval coverage
<code>na.rm</code>	set to TRUE to remove NAs before proceeding.

**Value**

a scalar with a value of 1.0 indicating symmetry

**Author(s)**

Frank Harrell

ExProb.blrm

*Function Generator for Exceedance Probabilities for `blrm()`***Description**

For a `blrm()` object generates a function for computing the estimates of the function  $\text{Prob}(Y \geq y)$  given one or more values of the linear predictor using the reference (median) intercept. This function can optionally be evaluated at only a set of user-specified  $y$  values, otherwise a right-step function is returned. There is a plot method for plotting the step functions, and if more than one linear predictor was evaluated multiple step functions are drawn. ExProb is especially useful for `nomogram()`. The linear predictor argument is a posterior summarized linear predictor `lp` (e.g. using posterior mean of intercepts and slopes) computed at the reference intercept. `lptau` must be provided when call the created function if the model is a partial proportional odds model.

**Usage**

```
## S3 method for class 'blrm'
ExProb(object, posterior.summary = c("mean", "median"), ...)
```

**Arguments**

<code>object</code>	a <code>blrm()</code> fit
<code>posterior.summary</code>	defaults to posterior mean; may also specify "median". Must be consistent with the summary used when creating <code>lp</code> .
<code>...</code>	ignored

**Value**

an R function

**Author(s)**

Frank Harrell

---

 getParamCoef

*Get a Bayesian Parameter Vector Summary*


---

**Description**

Retrieves posterior mean, median, or mode (if available)

**Usage**

```
getParamCoef(
  fit,
  posterior.summary = c("mean", "median", "mode"),
  what = c("both", "betas", "taus")
)
```

**Arguments**

`fit` a Bayesian model fit from `rmsb`

`posterior.summary` which summary statistic (Bayesian point estimate) to fetch

`what` specifies which coefficients to include. Default is all. Specify `what="betas"` to include only intercepts and betas if the model is a partial proportional odds model (i.e., exclude the tau parameters). Specify `what="taus"` to include only the tau parameters.

**Value**

vector of regression coefficients

**Author(s)**

Frank Harrell

---

 HPDint

*Highest Posterior Density Interval*


---

**Description**

Adapts code from `coda::HPDinterval()` to compute a highest posterior density interval from posterior samples for a single parameter. Quoting from the coda help file, for each parameter the interval is constructed from the empirical cdf of the sample as the shortest interval for which the difference in the ecdf values of the endpoints is the nominal probability. Assuming that the distribution is not severely multimodal, this is the HPD interval.

**Usage**

```
HPDint(x, prob = 0.95)
```

**Arguments**

x	a vector of posterior draws
prob	desired probability coverage

**Value**

a 2-vector with elements Lower and Upper

**Author(s)**

Douglas Bates and Frank Harrell

---

Mean.blrm

*Function Generator for Mean Y for `blrm()`*

---

**Description**

Creates a function to turn a posterior summarized linear predictor lp (e.g. using posterior mean of intercepts and slopes) computed at the reference intercept into e.g. an estimate of mean Y using the posterior mean of all the intercept. lptau must be provided when call the created function if the model is a partial proportional odds model.

**Usage**

```
## S3 method for class 'blrm'
Mean(object, codes = FALSE, posterior.summary = c("mean", "median"), ...)
```

**Arguments**

object	a <code>blrm()</code> fit
codes	if TRUE, use the integer codes 1, 2, ..., k for the k-level response in computing the predicted mean response.
posterior.summary	defaults to posterior mean; may also specify "median". Must be consistent with the summary used when creating lp.
...	ignored

**Value**

an R function

**Author(s)**

Frank Harrell

---

pdensityContour

*Bivariate Posterior Contour*


---

### Description

Computes coordinates of a highest density contour containing a given probability volume given a sample from a continuous bivariate distribution, and optionally plots. The default method assumes an elliptical shape, but one can optionally use a kernel density estimator. Code adapted from `embbook::HPDregionplot`. See <https://www.sumsar.net/blog/2014/11/how-to-summarize-a-2d-posterior-using>

### Usage

```
pdensityContour(
  x,
  y,
  method = c("ellipse", "kernel"),
  prob = 0.95,
  otherprob = c(0.01, 0.1, 0.25, 0.5, 0.75, 0.9),
  h = c(1.3 * MASS::bandwidth.nrd(x), 1.3 * MASS::bandwidth.nrd(y)),
  n = 70,
  pl = FALSE
)
```

### Arguments

<code>x</code>	a numeric vector
<code>y</code>	a numeric vector the same length of <code>x</code>
<code>method</code>	defaults to 'ellipse', can be set to 'kernel'
<code>prob</code>	main probability coverage (the only one for <code>method='ellipse'</code> )
<code>otherprob</code>	vector of other probability coverages for <code>method='kernel'</code>
<code>h</code>	vector of bandwidths for <code>x</code> and <code>y</code> . See <code>MASS::kde2d()</code> .
<code>n</code>	number of grid points in each direction, defaulting to normal reference bandwidth (see <code>bandwidth.nrd</code> ).
<code>pl</code>	set to <code>TRUE</code> to plot contours

### Value

a 2-column matrix with `x` and `y` coordinates unless `pl=TRUE` in which case a `ggplot2` graphic is returned

### Author(s)

Ben Bolker and Frank Harrell



---

`plot.PostF`*Plot Posterior Density of PostF*

---

### Description

Computes highest posterior density and posterior mean and median as vertical lines, and plots these on the density function. You can transform the posterior draws while plotting.

### Usage

```
## S3 method for class 'PostF'
plot(
  x,
  ...,
  cint = 0.95,
  label = NULL,
  type = c("linetype", "facet"),
  ltitle = ""
)
```

### Arguments

<code>x</code>	result of running a function created by <code>PostF</code>
<code>...</code>	other results created by such functions
<code>cint</code>	interval probability
<code>label</code>	x-axis label if not the expression originally evaluated. When more than one result is plotted, <code>label</code> is a vector of character strings, one for each result.
<code>type</code>	when plotting more than one result specifies whether to make one plot distinguishing results by line type, or whether to make separate panels
<code>ltitle</code>	used of <code>type='linetype'</code> to specify name of legend for the line types

### Value

ggplot2 object

### Author(s)

Frank Harrell

**Description**

For an rms Bayesian fit object, plots posterior densities for selected parameters along with posterior mode, mean, median, and highest posterior density interval. If the fit was produced by `stackMI` the density represents the distribution after stacking the posterior draws over imputations, and the per-imputation density is also drawn as pale curves. If exactly two parameters are being plotted and `bivar=TRUE`, highest bivariate posterior density contours are plotted instead, for a variety of prob values including the one specified, using

**Usage**

```
## S3 method for class 'rmsb'
plot(
  x,
  which = NULL,
  nrow = NULL,
  ncol = NULL,
  prob = 0.95,
  bivar = FALSE,
  bivarmethod = c("ellipse", "kernel"),
  ...
)
```

**Arguments**

<code>x</code>	an rms Bayesian fit object
<code>which</code>	names of parameters to plot, defaulting to all non-intercepts. Can instead be a vector of integers.
<code>nrow</code>	number of rows of plots
<code>ncol</code>	number of columns of plots
<code>prob</code>	probability for HPD interval
<code>bivar</code>	set to TRUE to plot bivariate density contours instead of univariate results (ignored if the number of parameters plotted is not exactly two)
<code>bivarmethod</code>	passed as method argument to <code>pdensityContour</code>
<code>...</code>	passed to <code>pdensityContour</code>

**Value**

ggplot2 object

**Author(s)**

Frank Harrell

**Description**

From a Bayesian fit object such as that from `blrm()` generates an R function for evaluating the probability that an assertion is true. The probability, within simulation error, is the proportion of times the assertion is true over the posterior draws. If the assertion does not evaluate to a logical or 0/1 quantity, it is taken as a continuous derived parameter and the vector of draws for that parameter is returned and can be passed to the PostF plot method. PostF can also be used on objects created by `contrast.rms`

**Usage**

```
PostF(fit, name = c("short", "orig"), pr = FALSE)
```

**Arguments**

<code>fit</code>	a Bayesian fit or <code>contrast.rms</code> object
<code>name</code>	specifies whether assertions will refer to shortened parameter names (the default) or original names. Shorted names are of the form <code>a1, ..., ak</code> where <code>k</code> is the number of intercepts in the model, and <code>b1, ..., bp</code> where <code>p</code> is the number of non-intercepts. When using original names that are not legal R variable names, you must enclose them in backticks. For contrast objects, <code>name</code> is ignored and you must use contrast names. The <code>cnames</code> argument to <code>contrast.rms</code> is handy for assigning your own names.
<code>pr</code>	set to TRUE to have a table of short names and original names printed when <code>name='short'</code> . For contrasts the contrast names are printed if <code>pr=TRUE</code> .

**Value**

an R function

**Author(s)**

Frank Harrell

**Examples**

```
## Not run:
f <- blrm(y ~ age + sex)
P <- PostF(f)
P(b2 > 0)      # Model is a1 + b1*age + b2*(sex == 'male')
P(b1 < 0 & b2 > 0) # Post prob of a compound assertion
# To compute probabilities using original parameter names:
P <- PostF(f, name='orig')
P(age < 0)     # Post prob of negative age effect
P(`sex=male` > 0)
```

```

f <- blrm(y ~ sex + pol(age, 2))
P <- PostF(f)
# Compute and plot posterior density of the vertex of the
# quadratic age effect
plot(P(-b2 / (2 * b3)))

# The following would be useful in age and sex interacted
k <- contrast(f, list(age=c(30, 50), sex='male'),
              list(age=c(30, 50), sex='female'),
              cnames=c('age 30 M-F', 'age 50 M-F'))
P <- PostF(k)
P(`age 30 M-F` > 0 & `age 50 M-F` > 0)
##'
## End(Not run)

```

---

predict.blrm

*Make predictions from a `blrm()` fit*

---

## Description

Predict method for `blrm()` objects

## Usage

```

## S3 method for class 'blrm'
predict(
  object,
  ...,
  kint = NULL,
  ycut = NULL,
  zcppo = TRUE,
  Zmatrix = TRUE,
  fun = NULL,
  funint = TRUE,
  type = c("lp", "fitted", "fitted.ind", "mean", "x", "data.frame", "terms", "cterm",
           "ccterm", "adjto", "adjto.data.frame", "model.frame"),
  se.fit = FALSE,
  codes = FALSE,
  posterior.summary = c("mean", "median", "all"),
  cint = 0.95
)

```

## Arguments

object, ..., type, se.fit, codes  
 see `rms::predict.lrm()`

kind	This is only useful in a multiple intercept model such as the ordinal logistic model. There to use to second of three intercepts, for example, specify kind=2. The default is the middle intercept corresponding to the median y. You can specify ycut instead, and the intercept corresponding to $Y \geq ycut$ will be used for kind.
ycut	for an ordinal model specifies the Y cutoff to use in evaluating departures from proportional odds, when the constrained partial proportional odds model is used. When omitted, ycut is implied by kind. The only time it is absolutely mandatory to specify ycut is when computing an effect (e.g., odds ratio) at a level of the response variable that did not occur in the data. This would only occur when the cppo function given to blrm is a continuous function. If type='x' and neither kind nor ycut are given, the partial PO part of the design matrix is not multiplied by the cppo function. If type='x', the number of predicted observations is 1, ycut is longer than 1, and zcppo is TRUE, predictions are duplicated to the length of ycut as it is assumed that the user wants to see the effect of varying ycut, e.g., to see cutoff-specific odds ratios.
zcppo	applies only to type='x' for a constrained partial PO model. Set to FALSE to prevent multiplication of Z matrix by cppo(ycut).
Zmatrix	set to FALSE to exclude the partial PO Z matrix from the returned design matrix if type='x'
fun	a function to evaluate on the linear predictor, e.g. a function created by <a href="#">Mean.blrm()</a> or <a href="#">Quantile.blrm()</a>
funint	set to FALSE if fun is not a function such as the result of <a href="#">Mean.blrm()</a> , <a href="#">Quantile.blrm()</a> , or <a href="#">ExProb.blrm()</a> that contains an intercepts argument
posterior.summary	set to 'median' or 'mode' to use posterior median/mode instead of mean. For some types set to 'all' to compute the needed quantity for all posterior draws, and return one more dimension in the array.
cint	probability for highest posterior density interval. Set to FALSE to suppress calculation of the interval.

**Value**

a data frame, matrix, or vector with posterior summaries for the requested quantity, plus an attribute 'draws' that has all the posterior draws for that quantity. For type='fitted' and type='fitted.ind' this attribute is a 3-dimensional array representing draws x observations generating predictions x levels of Y.

**Author(s)**

Frank Harrell

**See Also**

[rms::predict.lrm\(\)](#)

**Examples**

```
## Not run:
f <- blrm(...)
predict(f, newdata, type='...', posterior.summary='median')

## End(Not run)
```

---

```
print.blrm          Print blrm() Results
```

---

**Description**

Prints main results from `blrm()` along with indexes and predictive accuracy and their highest posterior density intervals computed from `blrmStats`.

**Usage**

```
## S3 method for class 'blrm'
print(
  x,
  dec = 4,
  coefs = TRUE,
  intercepts = x$non.slopes < 10,
  prob = 0.95,
  ns = 400,
  title = NULL,
  ...
)
```

**Arguments**

<code>x</code>	object created by <code>blrm()</code>
<code>dec</code>	number of digits to print to the right of the decimal
<code>coefs</code>	specify <code>FALSE</code> to suppress printing parameter estimates, and in integer <code>k</code> to print only the first <code>k</code>
<code>intercepts</code>	set to <code>FALSE</code> to suppress printing intercepts. Default is to print them unless there are more than 9.
<code>prob</code>	HPD interval probability for summary indexes
<code>ns</code>	number of random samples of the posterior draws for use in computing HPD intervals for accuracy indexes
<code>title</code>	title of output, constructed by default
<code>...</code>	passed to <code>prModFit</code>

**Author(s)**

Frank Harrell

**Examples**

```
## Not run:
f <- blrm(...)
options(lang='html') # default is lang='plain'; also can be latex
f # print using defaults
print(f, posterior.summary='median') # instead of post. means

## End(Not run)
```

---

print.blrmStats      *Print Details for blrmStats Predictive Accuracy Measures*

---

**Description**

Prints results of blrmStats with brief explanations

**Usage**

```
## S3 method for class 'blrmStats'
print(x, dec = 3, ...)
```

**Arguments**

x	an object produced by blrmStats
dec	number of digits to round indexes
...	ignored

**Author(s)**

Frank Harrell

**Examples**

```
## Not run:
f <- blrm(...)
s <- blrmStats(...)
s # print with defaults
print(s, dec=4)

## End(Not run)
```

---

```
print.predict.blrm    Print Predictions for blrm()
```

---

**Description**

Prints the summary portion of the results of `predict.blrm`

**Usage**

```
## S3 method for class 'predict.blrm'
print(x, digits = 3, ...)
```

**Arguments**

<code>x</code>	result from <code>predict.blrm</code>
<code>digits</code>	number of digits to round numeric results
<code>...</code>	ignored

**Author(s)**

Frank Harrell

---

```
print.rmsb    Basic Print for Bayesian Parameter Summary
```

---

**Description**

For a Bayesian regression fit prints the posterior mean, median, SE, highest posterior density interval, and symmetry coefficient from the posterior draws. For a given parameter, the symmetry measure is computed using the `distSym` function.

**Usage**

```
## S3 method for class 'rmsb'
print(x, prob = 0.95, dec = 4, intercepts = TRUE, pr = TRUE, ...)
```

**Arguments**

<code>x</code>	an object created by an rms Bayesian fitting function
<code>prob</code>	HPD interval coverage probability (default is 0.95)
<code>dec</code>	amount of rounding (digits to the right of the decimal)
<code>intercepts</code>	set to FALSE to not print intercepts
<code>pr</code>	set to FALSE to return an unrounded matrix and not print
<code>...</code>	ignored



**Value**

matrix (rounded if pr=TRUE)

**Author(s)**

Frank Harrell

**Examples**

```
## Not run:
f <- blrm(...)
print.rmsb(f)

## End(Not run)
```

---

Quantile.blrm

*Function Generator for Quantiles of Y for `blrm()`*

---

**Description**

Creates a function to turn a posterior summarized linear predictor  $lp$  (e.g. using posterior mean of intercepts and slopes) computed at the reference intercept into e.g. an estimate of a quantile of  $Y$  using the posterior mean of all the intercepts. `lptau` must be provided when call the created function if the model is a partial proportional odds model.

**Usage**

```
## S3 method for class 'blrm'
Quantile(object, codes = FALSE, posterior.summary = c("mean", "median"), ...)
```

**Arguments**

<code>object</code>	a <code>blrm()</code> fit
<code>codes</code>	if TRUE, use the integer codes $1, 2, \dots, k$ for the $k$ -level response in computing the quantile
<code>posterior.summary</code>	defaults to posterior mean; may also specify "median". Must be consistent with the summary used when creating $lp$ .
<code>...</code>	ignored

**Value**

an R function

**Author(s)**

Frank Harrell

---

`selectedQr`*QR Decomposition Preserving Selected Columns*

---

### Description

Runs a matrix through the QR decomposition and returns the transformed matrix and the forward and inverse transforming matrices R, Rinv. If columns of the input matrix X are centered the QR transformed matrix will be orthogonal. This is helpful in understanding the transformation and in scaling prior distributions on the transformed scale. `not` can be specified to keep selected columns as-is. `cornerQr` leaves the last column of X alone (possibly after centering). When `not` is specified, the square transforming matrices have appropriate identity submatrices inserted so that recreation of original X is automatic.

### Usage

```
selectedQr(X, not = NULL, corner = FALSE, center = TRUE)
```

### Arguments

<code>X</code>	a numeric matrix
<code>not</code>	an integer vector specifying which columns of X are to be kept with their original values
<code>corner</code>	set to FALSE to not treat the last column specially. You may not specify both <code>not</code> and <code>corner</code> .
<code>center</code>	set to FALSE to not center columns of X first

### Value

list with elements X, R, Rinv, xbar where xbar is the vector of means (vector of zeros if `center=FALSE`)

### Author(s)

Ben Goodrich and Frank Harrell

### Examples

```
x <- 1 : 10
X <- cbind(x, x^2)
w <- selectedQr(X)
w
with(w, X %**% R) # = scale(X, center=TRUE, scale=FALSE)
Xqr <- w$X
plot(X[, 1], Xqr[, 1])
plot(X[, 1], Xqr[, 2])
cov(X)
cov(Xqr)
```

```
X <- cbind(x, x^3, x^4, x^2)
w <- selectedQr(X, not=2:3)
with(w, X %>% R)
```

---

stackMI

*Bayesian Model Fitting and Stacking for Multiple Imputation*


---

## Description

Runs an rmsb package Bayesian fitting function such as `blrm` separately for each completed dataset given a multiple imputation result such as one produced by `Hmisc::aregImpute`. Stacks the posterior draws and diagnostics across all imputations, and computes parameter summaries on the stacked posterior draws.

## Usage

```
stackMI(
  formula,
  fitter,
  xtrans,
  data = NULL,
  n.impute = xtrans$n.impute,
  dtrans = NULL,
  derived = NULL,
  subset = NULL,
  refresh = 0,
  progress = if (refresh > 0) "stan-progress.txt" else "",
  file = NULL,
  ...
)
```

## Arguments

<code>formula</code>	a model formula
<code>fitter</code>	a Bayesian fitter
<code>xtrans</code>	an object created by <code>transcan</code> , <code>aregImpute</code> , or <code>mice</code>
<code>data</code>	data frame
<code>n.impute</code>	number of imputations to run, default is the number saved in <code>xtrans</code>
<code>dtrans</code>	see <code>Hmisc::fit.mult.impute</code>
<code>derived</code>	see <code>Hmisc::fit.mult.impute</code>
<code>subset</code>	an integer or logical vector specifying the subset of observations to fit
<code>refresh</code>	see <code>rstan::sampling</code> . The default is 0, indicating that no progress notes are output. If <code>refresh &gt; 0</code> and <code>progress</code> is not <code>' '</code> , progress output will be appended to file <code>progress</code> . The default file name is <code>'stan-progress.txt'</code> .

progress	see refresh. Defaults to '' if refresh = 0. Note: If running interactively but not under RStudio, rstan will open a browser window for monitoring progress.
file	optional file name in which to store results in RDS format. If file is given and it already exists, and none of the arguments to stackMI have changed since that fit, the fit object from file is immediately returned. So if the model, data, and imputations have not changed nothing needs to be computed.
...	arguments passed to fitter

**Value**

an rmsb fit object with expanded posterior draws and diagnostics

**Author(s)**

Frank Harrell

---

stanDx

*Print Stan Diagnostics*

---

**Description**

Retrieves the effect samples sizes and Rhats computed after a fitting function ran rstan, and prepares it for printing. If the fit was created by stackImpute, the diagnostics for all imputations are printed (separately).

**Usage**

```
stanDx(object)
```

**Arguments**

object	an object created by an rms package Bayesian fitting function such as <a href="#">blrm()</a> or <a href="#">stackMI()</a>
--------	---

**Value**

matrix suitable for printing

**Author(s)**

Frank Harrell

**Examples**

```
## Not run:
  f <- blrm(...)
  stanDx(f)

## End(Not run)
```

stanDxplot

*Diagnostic Trace Plots***Description**

For an rms Bayesian fit object, uses by default the stored posterior draws to check convergence properties of posterior sampling. If instead `rstan=TRUE`, calls the `rstan` traceplot function on the `rstan` object inside the `rmsb` object, to check properties of posterior sampling. If `rstan=TRUE` and the `rstan` object has been removed and `previous=TRUE`, attempts to find an already existing plot created by a previous run of the knitr chunk, assuming it was the `plotno` numbered plot of the chunk.

**Usage**

```
stanDxplot(
  x,
  which = NULL,
  rstan = FALSE,
  previous = TRUE,
  plotno = 1,
  rev = FALSE,
  stripsize = 8,
  ...
)
```

**Arguments**

<code>x</code>	an rms Bayesian fit object
<code>which</code>	names of parameters to plot, defaulting to all non-intercepts. When <code>rstan=FALSE</code> these are the friendly rms names, otherwise they are the <code>rstan</code> parameter names. If the model fit was run through <code>stackMI</code> for multiple imputation, the number of traces is multiplied by the number of imputations. Set to 'ALL' to plot all parameters.
<code>rstan</code>	set to TRUE to use <code>rstan::traceplot()</code> on a (presumed) stored <code>rstan</code> object in <code>x</code> , otherwise only real iterations are plotted and parameter values are shown as points instead of lines, with chains separated
<code>previous</code>	see details
<code>plotno</code>	see details
<code>rev</code>	set to TRUE to reverse direction for faceting chains
<code>stripsize</code>	specifies size of chain facet label text, default is 8
<code>...</code>	passed to <code>rstan::traceplot()</code>

**Value**

ggplot2 object if `rstan` object was in `x`

**Author(s)**

Frank Harrell

---

stanGet

*Get Stan Output*

---

**Description**

Extracts the object created by `rstan::sampling()` so that standard Stan diagnostics can be run from it

**Usage**

```
stanGet(object)
```

**Arguments**

`object` an object created by an rms package Bayesian fitting function

**Value**

the object created by `rstan::sampling()`

**Author(s)**

Frank Harrell

**Examples**

```
## Not run:  
f <- blrm(...)  
s <- stanGet(f)  
  
## End(Not run)
```

---

tauFetch

*Fetch Partial Proportional Odds Parameters*


---

**Description**

Fetches matrix of posterior draws for partial proportional odds parameters (taus) for a given intercept. Can also form a matrix containing both regular parameters and taus, or for just non-taus. For the constrained partial proportional odds model the function returns the appropriate cppo function value multiplied by tau (tau being a vector in this case and not a matrix).

**Usage**

```
tauFetch(fit, intercept, what = c("tau", "nontau", "both"))
```

**Arguments**

fit	an object created by <code>blrm()</code>
intercept	integer specifying which intercept to fetch
what	specifies the result to return

**Value**

matrix with number of rows equal to the number of original draws

**Author(s)**

Frank Harrell

---

vcov.rmsb

*Variance-Covariance Matrix*


---

**Description**

Computes the variance-covariance matrix from the posterior draws by compute the sample covariance matrix of the draws

**Usage**

```
## S3 method for class 'rmsb'
vcov(object, regcoef.only = TRUE, intercepts = "all", ...)
```

**Arguments**

object	an object produced by an rms package Bayesian fitting function
regcoef.only	set to FALSE to also include non-regression coefficients such as shape/scale parameters
intercepts	set to 'all' to include all intercepts (the default), 'none' to exclude them all, or a vector of integers to get selected intercepts
...	ignored

**Value**

matrix

**Author(s)**

Frank Harrell

**See Also**

[rms::vcov.rms\(\)](#)

**Examples**

```
## Not run:  
f <- blrm(...)  
v <- vcov(f)  
  
## End(Not run)
```



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