

hyperSpec Plotting functions

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Reproducing the Examples in this Vignette

All spectra used in this manual are installed automatically with *hyperSpec*.

Note that some definitions are executed in `vignette.defs`, and others invisibly at the beginning of the file in order to have the code as similar as possible to interactive sessions.

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Suggested Packages

latticeExtra: available

deldir: available

rgl: available

ggplot2: available

In addition *tripack*, and *latticeist* are mentioned, but not used in this vignette.

Preliminary Calculations

For some plots of the `chondro` dataset, the pre-processed spectra and their cluster averages \pm one standard deviation are more suitable:

```

> chondro.preproc <- chondro - spc.fit.poly.below (chondro)
Fitting with npts.min = 15

> chondro.preproc <- chondro.preproc / rowMeans (chondro)
> chondro.preproc <- chondro.preproc - quantile (chondro, 0.05)
> cluster.cols <- c ("dark blue", "orange", "#C02020")
> cluster.meansd <- aggregate (chondro.preproc, chondro$clusters, mean_pm_sd)
> cluster.means <- aggregate (chondro.preproc, chondro$clusters, mean)

```

For details about the pre-processing, please refer to the example work flow in vignette ("chondro"), or the help ? chondro.

1 Predefined functions

hyperSpec comes with 6 major predefined plotting functions.

`plot` main switchyard for most plotting tasks

`levelplot` *hyperSpec* has a method for `lattice[?]` function `levelplot`

plotspc plots spectra

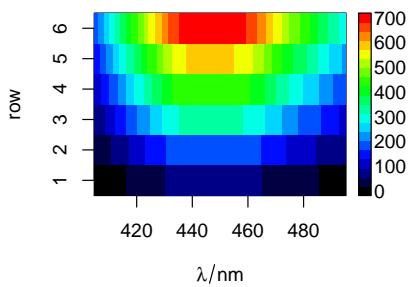
plotmat plots the spectra matrix

plotc calibration plot, time series, depth profile
plotc is a *lattice* function

plotmap more specialized version of **levelplot** for map or image plots.
plotmap is a *lattice* function

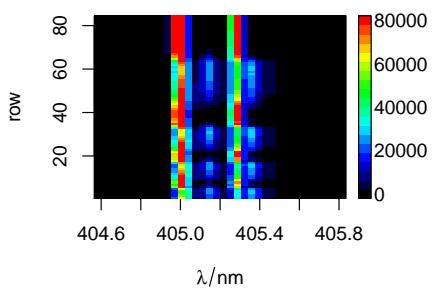
`plotvoronoi` more specialized version of `plotmap` that produces Voronoi tessellations.
`plotvoronoi` is a *lattice* function

`plotmap`, `plotvoronoi`, and `levelplot` are *lattice* functions. Therefore, in loops, functions, Sweave chunks, etc. the lattice object needs to be printed explicitly by e.g. `print(plotmap(object))` ([R FAQ: Why do lattice/trellis graphics not work?](#)).

plotspc

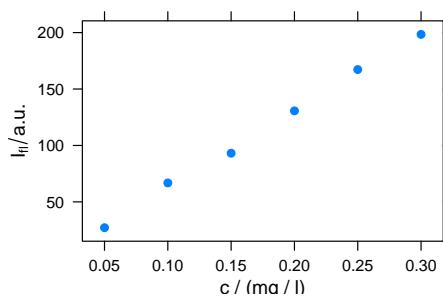
plots the spectra, i.e. the intensities \$spc over the wavelengths @wavelength.

> plotspc (flu)

plotmat

plots the spectra, i.e. the colour coded intensities \$spc over the wavelengths @wavelength and the row number.

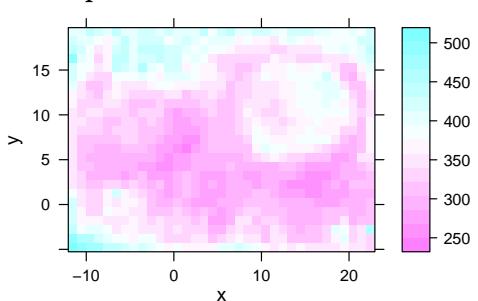
> plotmat (flu)

plotc

plots an intensity over a single other data column, e.g.

- calibration
- time series
- depth profile

> plotc (flu)

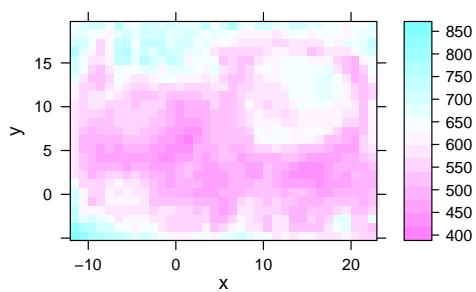
levelplot

plots a false colour map, defined by a formula.

> levelplot (spc ~ x * y, chondro, aspect = "iso")

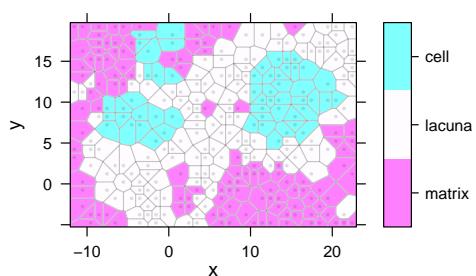
Warning: Only first wavelength is used for plotting

plotmap



plotmap is a specialized version of levelplot. It uses a single value (e.g. average intensity or cluster membership) over two data columns (default \$x and \$y)
> `plotmap (chondro)`

plotvoronoi

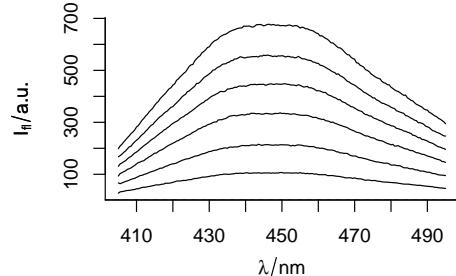


plotmap is a specialized version of levelplot. It uses a single value (e.g. average intensity or cluster membership) over two data columns (default \$x and \$y)
> `plotvoronoi (sample (chondro, 300), clusters ~ x * y)`

2 Arguments for plot

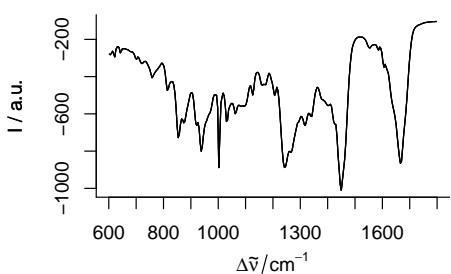
hyperSpec's `plot` method uses its second argument to determine which of the specialized plots to produce. This allows some handy abbreviations. All further arguments are handed over to the function actually producing the plot.

plot (x, "spc")



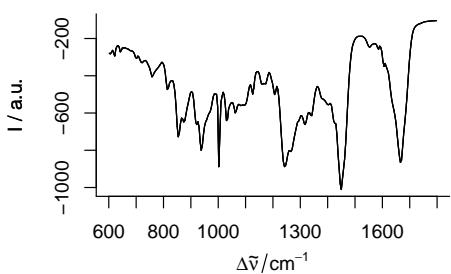
is equivalent to `plotspc (flu)`
> `plot (flu, "spc")`

```
plot (x, "spcmeansd")
```



plots mean spectrum ± 1 standard deviation
> plot (chondro.preproc, "spcmeansd")

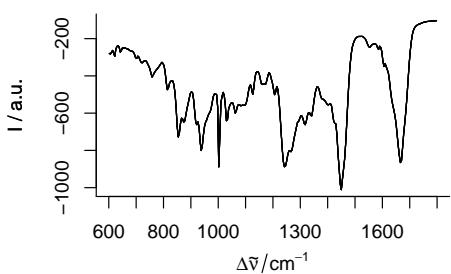
```
plot (x, "spcprctile")
```



plots median, 16th and 84th percentile for each wavelength.
For Gaussian distributed data, 16th, 50th and 84th percentile
are equal to mean \pm standard deviation. Spectroscopic data
frequently are not Gaussian distributed. The percentiles give
a better idea of the true distribution. They are also less
sensitive to outliers.

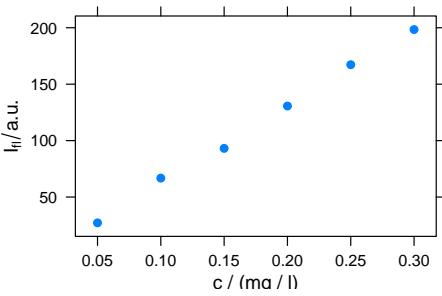
> plot (chondro.preproc, "spcprctile")

```
plot (x, "spcprctl5")
```



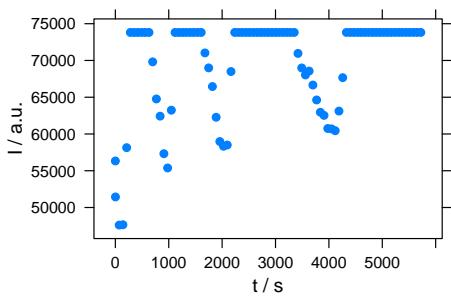
like "spcprctl" plus 5th and 95th percentile.
> plot (chondro.preproc, "spcprctl5")

```
plot (x, "c")
```



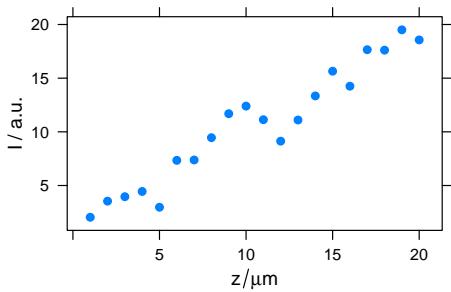
> plot (flu, "c")
is equivalent to plotc (flu)

plot (x, "ts")



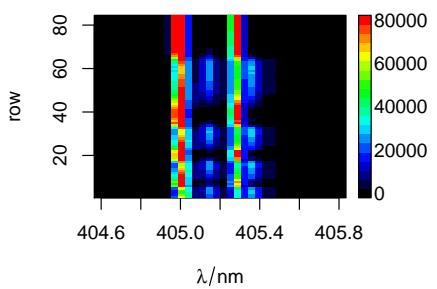
plots a time series plot
> plot (laser [,, 405], "ts")
equivalent to plotc (laser, spc ~ t)

plot (x, "depth")



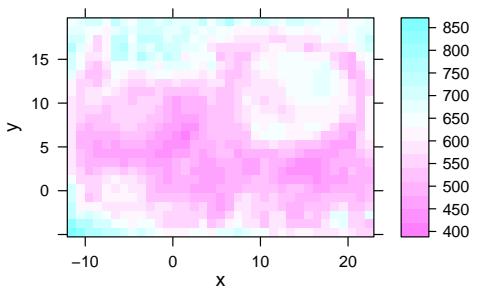
plots a depth profile plot
> depth.profile <- new ("hyperSpec",
+ spc = as.matrix (rnorm (20) + 1:20),
+ data = data.frame (z = 1 : 20),
+ labels = list (spc = "I / a.u.",
+ z = expression ('/'~(z, mu*m)),
+ .wavelength = expression (lambda)))
> plot (depth.profile, "depth")
the same as plotc (laser, spc ~ z)

plot (x, "mat")



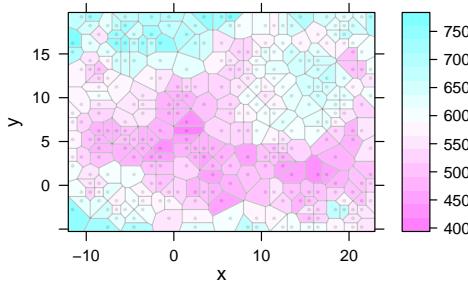
plots the spectra matrix.
> plot (laser, "mat")
Equivalent to
> plotmat (laser)
A lattice alternative is:
> levelplot (spc ~ .wavelength * .row, laser)

plot (x, "map")



is equivalent to plotmap (chondro)
> plot (chondro, "map")

```
plot (x, "voronoi")
```



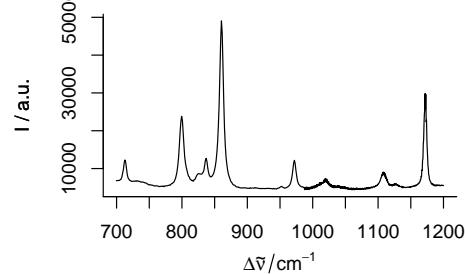
```
> plot (sample (chondro, 300), "voronoi")
See plotvoronoi
```

3 Spectra

plotspc

plotspc offers a variety of parameters for customized plots. To plot ...

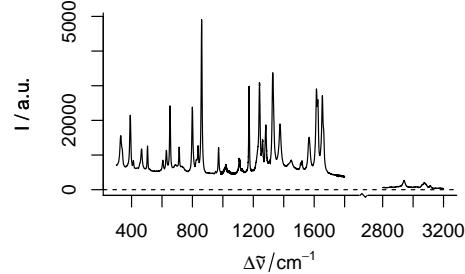
particular wavelength range



if only one wavelength range is needed, the extract command (see vignette ("introduction")) is handiest:

```
> plotspc (paracetamol [,, 700 ~ 1200])
wavelengths. If wl.range already contains indices use
wl.index = TRUE.
```

more wavelength ranges

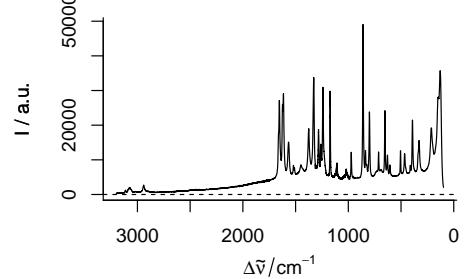


use wl.range = list (600 ~ 1800, 2800 ~ 3100). Cut the wavelength axis appropriately with xoffset = 750

```
> plotspc (paracetamol,
+           wl.range = c (300 ~ 1800, 2800 ~ max),
+           xoffset = 750)
```

If available, the package *plotrix*[1] is used to produce the cut mark.

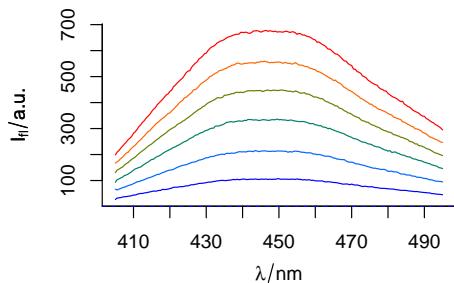
with reversed abscissa



use wl.reverse = TRUE

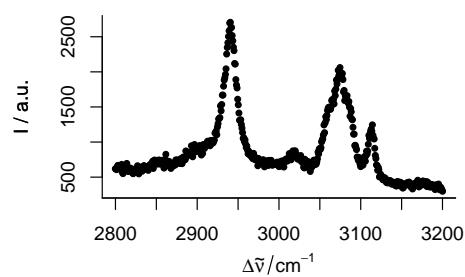
```
> plotspc (paracetamol, wl.reverse = TRUE )
```

in different colours



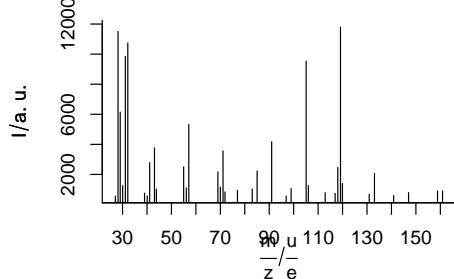
```
use col = vector.of.colours  
> plotspc (flu, col = matlab.dark.palette (6))
```

dots instead of lines



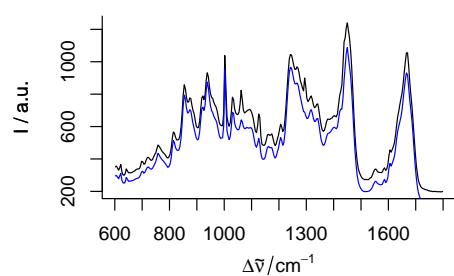
```
use lines.args = list (pch = 20, type = "p")  
> plotspc (paracetamol [,, 2800 ~ 3200],  
+           lines.args = list (pch = 20, type = "p"))
```

mass spectra



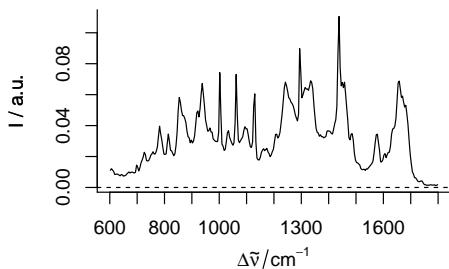
```
use lines.args = list (type = "h")  
> plot (barbiturates [[1]], lines.args = list (type = "h"))
```

more spectra into an existing plot



```
use add = TRUE  
> plotspc (chondro [ 30,,])  
> plotspc (chondro [300,,], add = TRUE, col = "blue")
```

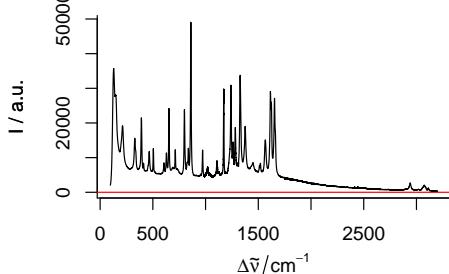
Summary characteristics



`func` may be used to calculate summary characteristics prior to plotting. To plot e.g. the standard deviation of the spectra, use:

```
> plotspc (chondro.preproc, func = sd)
```

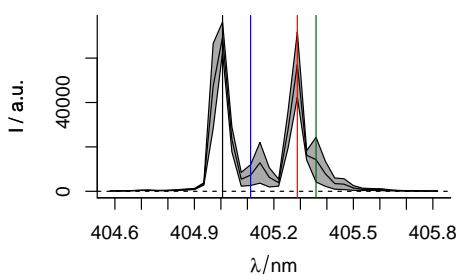
with different line at $I = 0$



`zeroline` takes a list with parameters to `abline`, NA suppresses the line.

```
> plotspc (paracetamol,
+           zeroline = list (col = "red"))
```

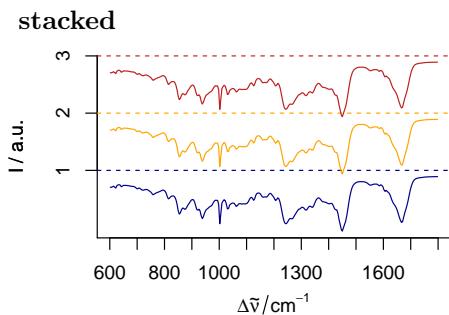
adding to a spectra plot



`plotspc` uses base graphics. After plotting the spectra, more content may be added to the graphic by `abline`, `lines`, `points`, etc.

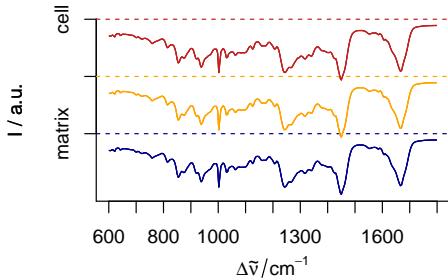
```
> plot (laser, "spcmeansd")
> abline (v = c(405.0063, 405.1121, 405.2885, 405.3591),
+          col = c("black", "blue", "red", "darkgreen"))
```

3.1 Stacked spectra



```
use stacked = TRUE
> plotspc (cluster.means,
+           col = cluster.cols,
+           stacked = TRUE)
```

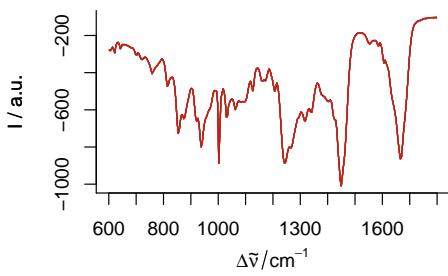
Stacking groups of spectra



The spectra to be stacked can be grouped: `stacked = factor`. Alternatively, the name of the grouping extra data column can be used:

```
> plot (cluster.meansd,
+       stacked = ".aggregate",
+       fill = ".aggregate",
+       col = cluster.cols)
```

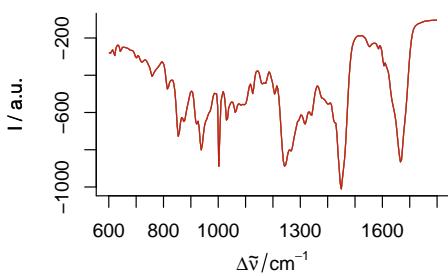
Manually giving yoffset



Stacking values can also be given manually as numeric values in `yoffset`:

```
> plotspc (cluster.meansd,
+            yoffset = rep (0:2, each = 3),
+            col = rep (cluster.cols, each = 3))
```

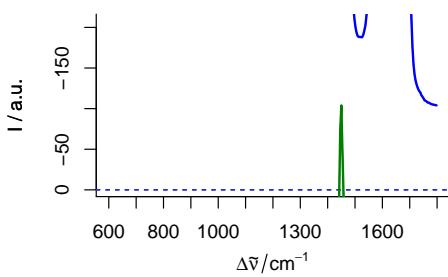
Dense stacking



To obtain a denser stacking:

```
> yoffsets <- apply (cluster.means [[], 2, diff)
> yoffsets <- - apply (yoffsets, 1, min)
> plot (cluster.means, yoffset = c (0, cumsum (yoffsets)),
+       col = cluster.cols)
```

Elaborate example



```
> yoffset <- apply (chondro.preproc, 2, quantile, c(0.05, 0.95))
> yoffset <- range (yoffset)
> plot(chondro.preproc[1],
+       plot.args = list (ylim = c (0, 2) * yoffset),
+       lines.args = list( type = "n"))
> yoffset <- (0:1) * diff (yoffset)
> for (i in 1 : 3){
+   plot(chondro.preproc, "spcprctl5", yoffset = yoffset [i],
+         col = "gray", add = TRUE)
+   plot (chondro.preproc [i], yoffset = yoffset [i],
+         col = matlab.dark.palette (3) [i], add = TRUE,
+         lines.args = list (lwd = 2))
+ }
```

`plotspc` allows fine grained customization of almost all aspects of the plot. This is possible by

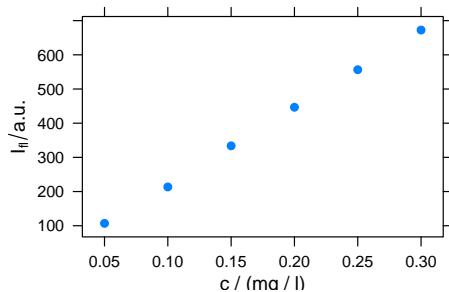
giving arguments to the functions that actually perform the plotting `plot` for setting up the plot area, `lines` for the plotting of the lines, `axis` for the axes, etc. The arguments for these functions should be given in lists as `plot.args`, `lines.args`, `axis.args`, etc.

4 Calibration Plots, (Depth) Profiles, and Time Series Plots

`plotc`

4.1 Calibration plots

Intensities over concentration

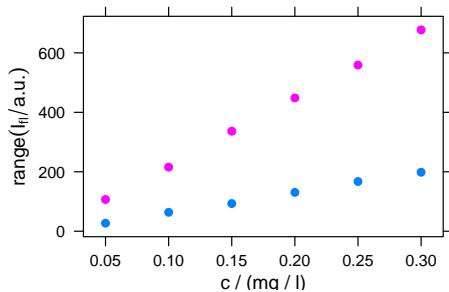


Plotting the Intensities of one wavelength over the concentration for univariate calibration:

> `plotc (flu [,, 450])`

The default is to use the first intensity only.

Summary Intensities over concentration

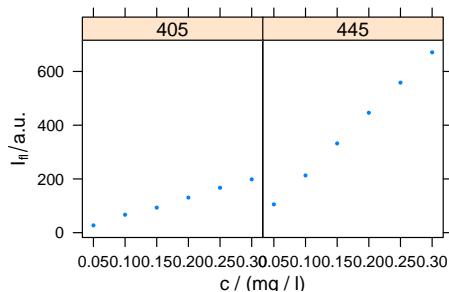


A function to compute a summary of the intensities before drawing can be used:

> `plotc (flu, func = range, groups = .wavelength)`

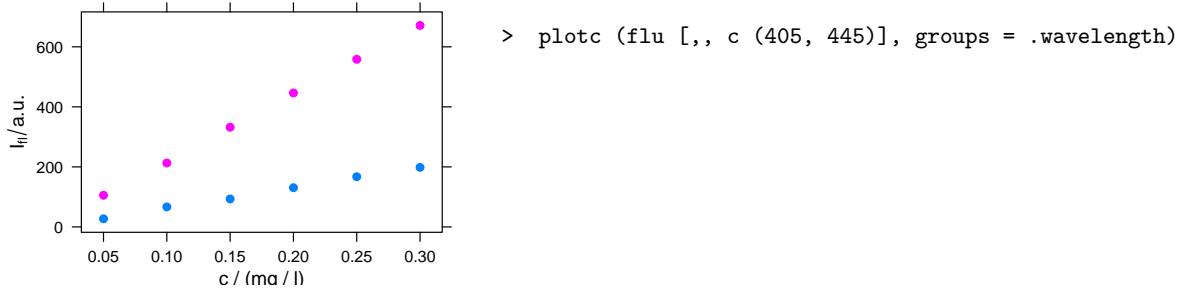
If `func` returns more than one value, the different results are accessible by `.wavelength`.

Conditioning: plotting more traces separately

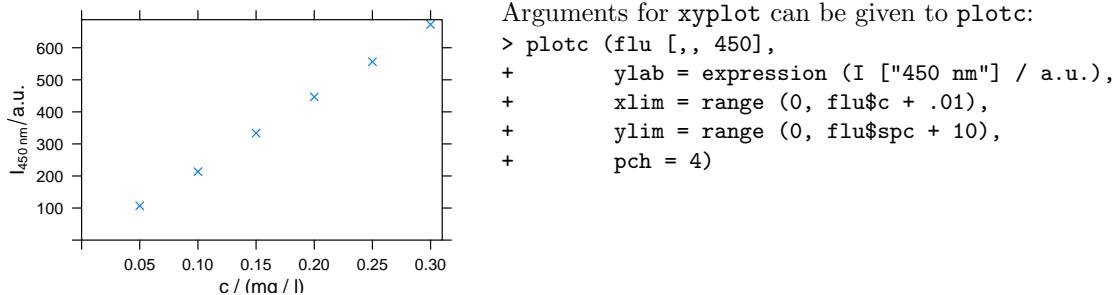


```
> plotc (flu [,, c (405, 445)], spc ~ c | .wavelength,
+           cex = .3, scales = list (alternating = c(1, 1)))
```

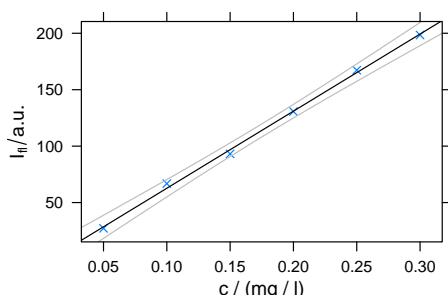
Grouping: plot more traces in one panel



Changing Axis Labels (and other parameters)



Adding things to the plot: customized panel function

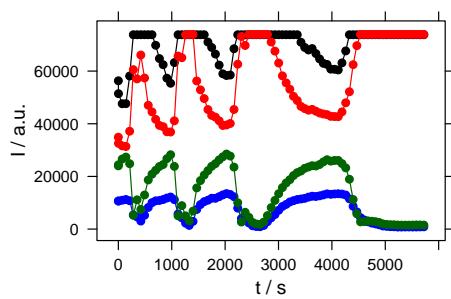


As `plotc` uses the *lattice* function `xyplot`, additions to the plot must be made via the panel function:

```
> panelcalibration <- function(x, y, ..., clim = range(x), level = .99)
+   panel.xyplot(x, y, ...)
+   lm <- lm(y ~ x)
+   panel.abline(coef(lm), ...)
+   cx <- seq(clim[1], clim[2], length.out = 50)
+   cy <- predict(lm, data.frame(x = cx),
+                 interval = "confidence",
+                 level = level)
+   panel.lines(cx, cy [,2], col = "gray")
+   panel.lines(cx, cy [,3], col = "gray")
+ }
> plotc(flu [,,405], panel = panelcalibration,
+         pch = 4, clim = c(0, 0.35), level = .99)
```

4.2 Time series and other Plots of the Type Intensity-over-Something

Abscissae other than c



Other abscissae may be specified by explicitly giving the model formula:

```
> plotc (laser [,, c(405.0063, 405.1121, 405.2885, 405.3591)],
+         spc ~ t,
+         groups = .wavelength,
+         type = "b",
+         col = c ("black", "blue", "red", "darkgreen"))
```

5 Levelplot

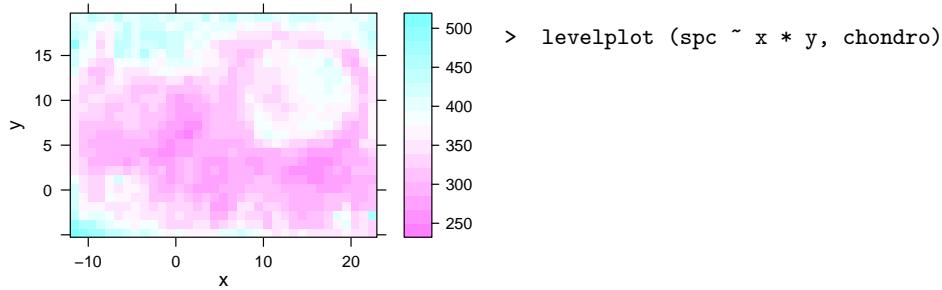
hyperSpec's *levelplot* can use two special column names:

.wavelength for the wavelengths

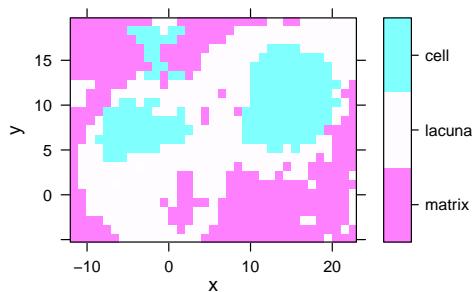
.row for the row index (i.e. spectrum number) in the data

Besides that, it behaves exactly like *levelplot*. Particularly, the data is given as the second argument:

levelplot



factors as z



If the colour-coded value is a factor, the display is adjusted to this fact:

```
> levelplot (clusters ~ x * y, chondro)
```

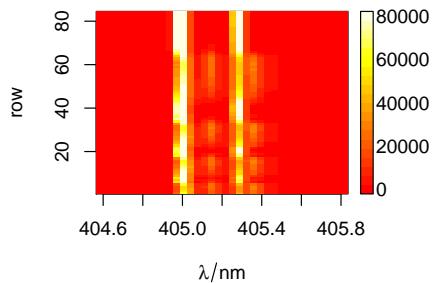
6 Spectra Matrix

It is often useful to plot the spectra against an additional coordinate, e.g. the time for time series, the depth for depth profiles, etc.

This can be done by `plot (object, "mat")`. The actual plotting is done by `image`, but `levelplot`

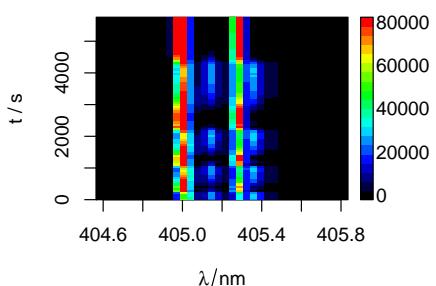
can produce spectra matrix plots as well and these plots can be grouped or conditioned.

different palette



```
> plot(laser, "mat", col = heat.colors(20))
is the same as
> plotmat(laser, col = heat.colors(20))
```

different y axis

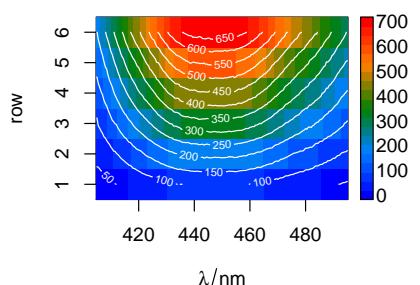


Using a different extra data column for the y axis:

```
> plotmat(laser, y = "t")
alternatively, y values and axis label can be given separately.

> plotmat(laser, y = laser$t, ylab = labels(laser, "t"))
```

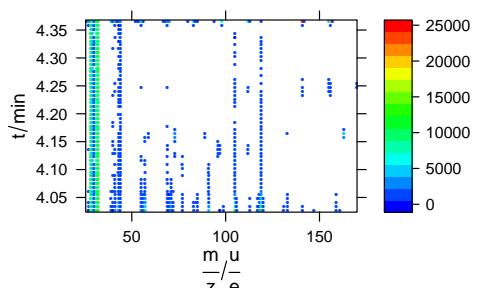
contour lines



Contour lines may be added:

```
> plotmat(flu, col = matlab.dark.palette(20))
> plotmat(flu, col = "white",
+           contour = TRUE, add = TRUE)
```

colour-coded points: levelplot with special panel function

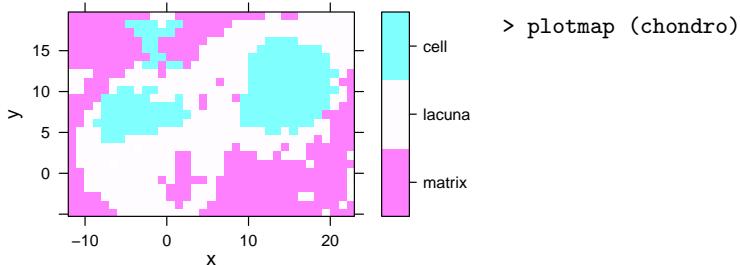


```
> require("latticeExtra")
> barb <- do.call(collapse, barbiturates[1:50])
> barb <- orderwl(barb)
> levelplot(spc ~ .wavelength * z, barb,
+            panel = panel.levelplot.points,
+            cex = .33, col.symbol = NA,
+            col.regions = matlab.palette)
```

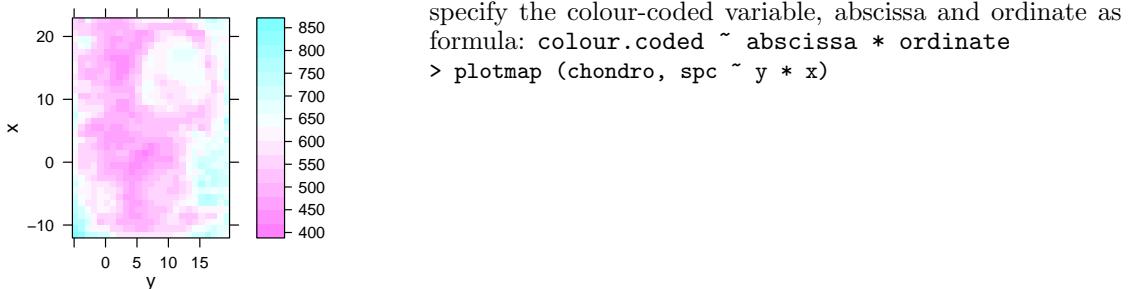
7 False-Colour Maps: plotmap

plotmap is a specialized version of levelplot. The spectral intensities may be summarized by a function before plotting (default: `mean`). The same scale is used for x and y axes (`aspect = "iso"`).

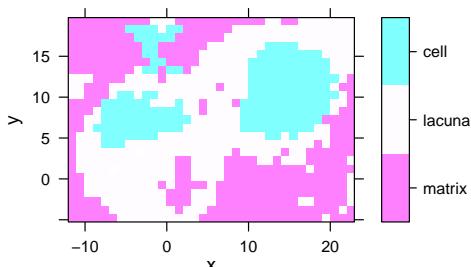
plotting map



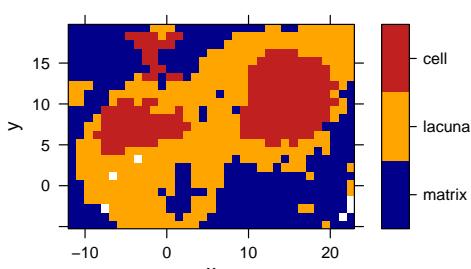
plotting maps with other than x and y



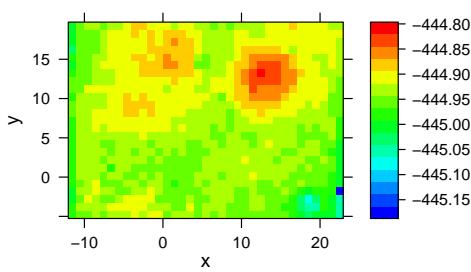
colour-coded factors



different palette



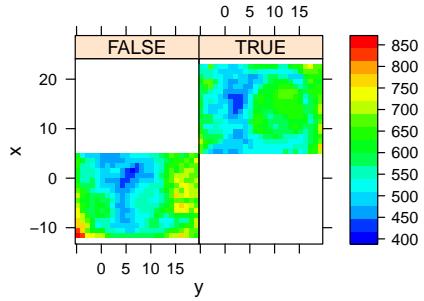
defined wavelengths



To plot a map of the average intensity at particular wavelengths use extraction:

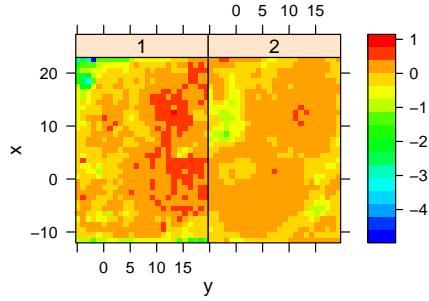
```
> plotmap (chondro.preproc [ , , c(728, 782, 1098,
+                               1240, 1482, 1577)],
+           col.regions = matlab.palette)
```

Conditioning



```
> plotmap (chondro,
+           spc ~ y * x | x > 5,
+           col.regions = matlab.palette(20))
```

Conditioning on .wavelength

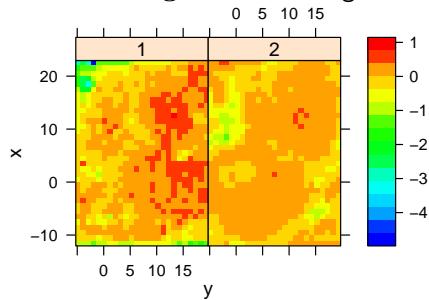


`plotmap` automatically applies the function in `func` before plotting. This defaults to the `mean`. In order to suppress this, use `func = NULL`. This allows conditioning on the wavelengths.

To plot e.g. the first two score maps of a principal component analysis:

```
> pca <- prcomp (~ spc, data = chondro.preproc$.)
> scores <- decomposition (chondro, pca$x,
+                           label.wavelength = "PC",
+                           label.spc = "score / a.u.")
> plotmap (scores [,,1:2],
+           spc ~ y * x | as.factor(.wavelength),
+           func = NULL,
+           col.regions = matlab.palette(20))
```

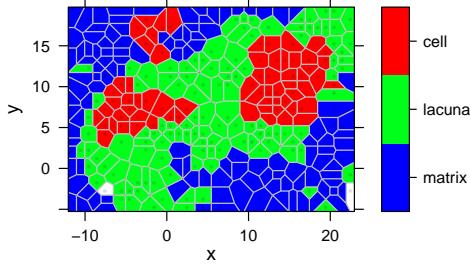
Conditioning on .wavelength II



Alternatively, use `levelplot` directly:

```
> levelplot (spc ~ y * x | as.factor(.wavelength),
+             scores [,,1:2],
+             aspect = "iso",
+             col.regions = matlab.palette(20))
```

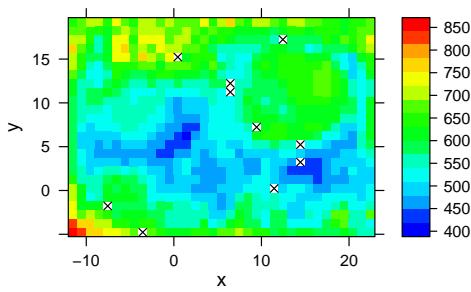
Voronoi plot



```
> plotvoronoi (sample (chondro, 300), clusters ~ x * y,
+               col.regions = matlab.palette(20))
```

Voronoi uses `panel.voronoi` from *latticeExtra*[2]. The tesselation is calculated by default using `deldir`[3], but `tripack`[4] can also be used. `tripack` seems to faster in general, but may “hang” with certain data sets (particularly regular grids with missing spectra as in this example). Furthermore, it is not FOSS (free and open source software), so users are kindly asked to review `tripack`’s license before using it.

Mark missing spectra

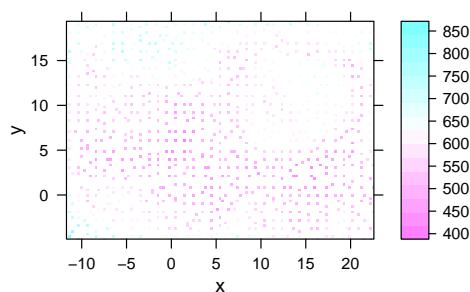


If the spectra come from a rectangular grid, missing positions can be marked with this panel function:

```
> mark.missing <- function (x, y, z, ...){
+   panel.levelplot (x, y, z, ...)
+
+   miss <- expand.grid (x = unique (x), y = unique (y))
+   miss <- merge (miss, data.frame (x, y, TRUE),
+                  all.x = TRUE)
+   miss <- miss [is.na (miss[, 3]),]
+   panel.xyplot (miss [, 1], miss [, 2], pch = 4, ...)
+ }
> plotmap (sample (chondro, 865),
+            col.regions = matlab.palette(20),
+            col = "black",
+            panel = mark.missing)
```

Unevenly spaced measurement grid

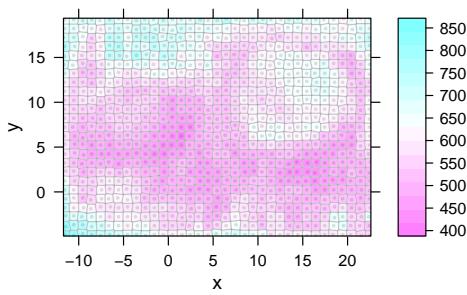
The panel function used by `plotmap` defaults to `panel.levelplot.raster` which assumes an evenly spaced measurement grid. Even if the spectra are measured on a nominally evenly spaced grid, the actual stage position may be slightly varying due to positioning inaccuracy and some manufacturers (e.g. Kaiser) record the position reported by the stage rather than the position requested by the stage control.



This leads to weird looking output with holes, and possibly wrong columns:

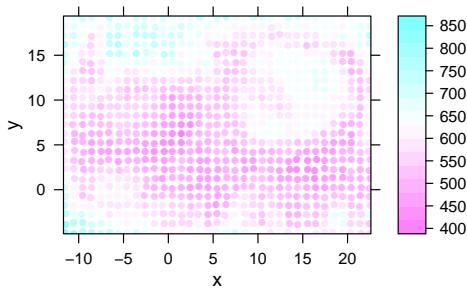
```
> plotmap (uneven)
```

The symptom of this situation are warnings about values in `x` and/or `y` not being equispaced; and that the output therefore may be wrong.



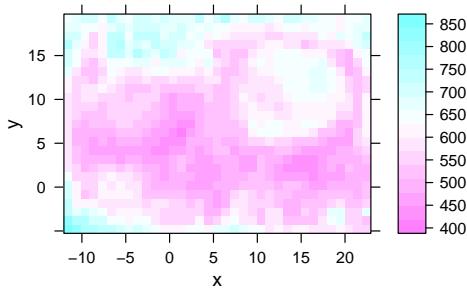
One possibility to obtain a correct map is using `plotvoronoi` instead which will construct a mosaic-like image with the respective “pixel” areas being centered around the actually recorded \$x and \$y position:

```
> plotvoronoi (uneven)
```



Another possibility that underlines a point shape of the measurements is switching to `latticeExtra::panel.levelplot.points`:

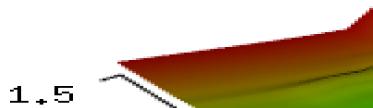
```
> plotmap (uneven, panel = panel.levelplot.points,
+           cex = 0.75, col.symbol=NA)
```



Alternatively, the measurement raster positions can be rounded to their nominal raster, e.g.:

```
> rx <- makeraster (uneven$x, start = -11.55, d = 1, tol = 0.3)
> uneven$x <- rx$x
> ry <- makeraster (uneven$y, start = -4.77, d = 1, tol = 0.3)
> uneven$y <- ry$x
> plotmap (uneven)
```

8 3D plots (with rgl)



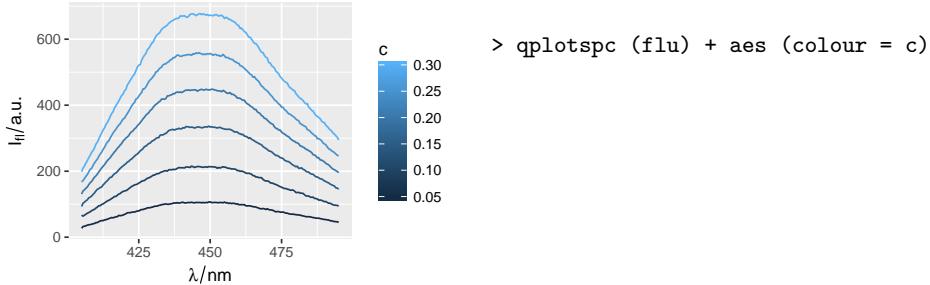
`rgl[5]` offers fast 3d plotting in R. As `rgl`'s axis annotations are sometimes awkward, they may better be set manually:

```
> laser <- laser [,,404.8 ~ 405.6] / 10000
> laser$t <- laser$t / 3600
> cols <- rep (matlab.palette (nrow (laser)), nwl (laser))
> surface3d (y = wl (laser), x = laser$t,
+             z = laser$spc, col = cols)
> aspect3d (c (1, 1, 0.25))
> axes3d (c ('x--', 'y--', 'z--'))
> axes3d ('y--', nticks = 25, labels= FALSE)
> mtext3d ("t / h", 'x--', line = 2.5)
> mtext3d ("lambda / nm", 'y--', line = 2.5)
> mtext3d ("I / a.u.", edge = 'z--', line = 2.5)
```

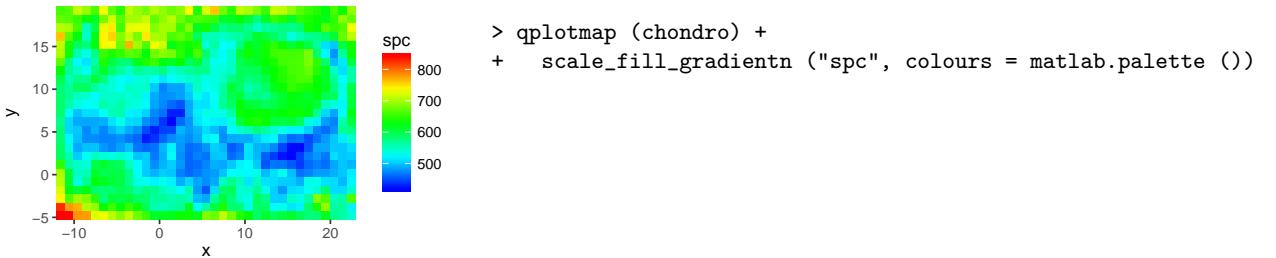
9 Using `ggplot2` with `hyperSpec` objects

`hyperSpec` objects do not yet directly support plotting with `ggplot2` [6]. Nevertheless, `ggplot2` graphics can easily be obtained, and `qplot*` equivalents to `plotspc` and `plotmap` are defined:

plot spectra with `as.long.df`



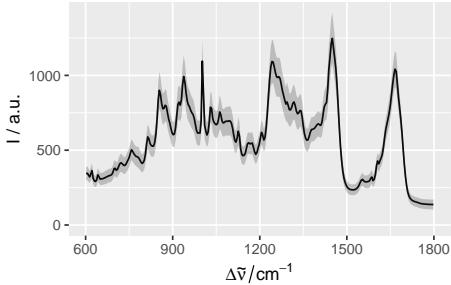
Map with `ggplot2`



The two special columns `.wavelength` and `.rownames` contain the wavelength axis and allow to distinguish the spectra.

For more general plotting, `as.long.df` transforms a `hyperSpec` object into a long-form `data.frame` that is suitable for `qplot`, while `as.t.df` produces a `data.frame` where each spectrum is one column, and an additional first column gives the wavelength (see “plotting mean \pm sd” below for an example). Long `data.frames` can be *very* memory consuming as they are of size $nrow \cdot nwl \times (ncol + 2)$ with respect to the dimensions of the `hyperSpec` object. Thus, e.g. the `chondro` data set (2 MB) as `hyperSpec` object) needs 24 MB as long-format `data.frame`. It is therefore highly recommended to calculate the particular data to be plotted beforehand.

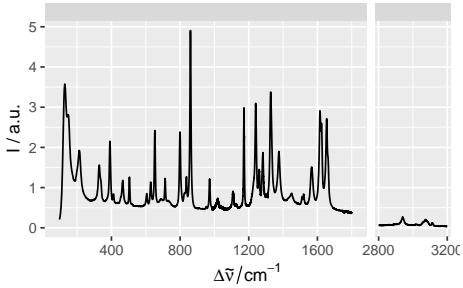
Mean \pm standard deviation with *ggplot2*



```
> qplotspc (mean (chondro)) +
+   geom_ribbon (aes (ymin = mean + sd,
+                     ymax = mean - sd,
+                     y = 0, group = NA),
+                     alpha = 0.25,
+                     data = as.t.df (mean_sd (chondro)))
```

Note that `qplotspc` specifies aesthetics `y = spc` and `groups = .rownames`, which do not have corresponding columns in the `data.frame` returned by `as.t.df`. These aesthetics must therefore be set manually in the aesthetics definition in `geom_ribbon` (or any other `geom_` that uses `as.t.df`). Otherwise, errors occur that object `spc` (and/or `.rownames`) cannot be found.

Cut spectra with *ggplot2*



Cut axes can be implemented by facetting:

```
> qplotspc (paracetamol / 1e4,
+            wl.range = c( min ~ 1800, 2800 ~ max)) +
+            scale_x_continuous (breaks = seq (0, 3200, 400))
```

10 Troubleshooting

10.1 No output is produced

`plotmap`, `plotvoronoi`, `levelplot`, and `plotc` use *lattice* functions. Therefore, in loops, functions, Sweave chunks, etc. the *lattice* object needs to be printed explicitly by `print (plotmap (object))` ([R FAQ: Why do lattice/trellis graphics not work?](#)). The same holds for *ggplot2* graphics.

For suggestions how the *lattice* functions can be redefined so that the result is printed without external `print` command, see the file `vignettes.defs`.

11 Interactive Graphics

hyperSpec offers basic interaction, `spc.identify` for spectra plots, and `map.identify` and `map.sel.poly` for maps. The first two identify points in spectra plots and map plots, respectively. `map.sel.poly` selects the part of a *hyperSpec* object that lies inside the user defined polygon.

11.1 `spc.identify`: finding out wavelength, intensity and spectrum

`spc.identify` allows to measure points in graphics produced by `plotspc`. It works correctly with reversed and cut wavelength axes.

```
> spc.identify (plotspc (paracetamol, wl.range = c (600 ~ 1800, 2800 ~ 3200), xoffset = 800))
```

The result is a `data.frame` with the indices of the spectra, the wavelength, and its intensity.

11.2 map.identify: finding a spectrum in a map plot

`map.identify` returns the spectra indices of the clicked points.

```
> map.identify (chondro)
```

11.3 map.sel.poly: selecting spectra inside a polygon in a map plot

`map.sel.poly` returns a logical indicating which spectra are inside the polygon drawn by the user:

```
> map.sel.poly (chondro)
```

11.4 Related functions provided by base graphics and lattice

For base graphics (as produced by `plotspc`), `locator` may be useful as well. It returns the clicked coordinates. Note that these are *not* transformed according to `xoffset` & Co.

For lattice graphics, `grid.locator` may be used instead. If it is not called in the panel function, a preceding call to `trellis.focus` is needed:

```
> plot (laser, "mat")
> trellis.focus ()
> grid.locator ()
```

`identify` (or `panel.identify` for lattice graphics) allows to identify points of the plot directly. Note that the returned indices correspond to the plotted object.

References

- [1] Lemon J. Plotrix: a package in the red light district of r. *R-News*, 6(4):8–12, 2006.
- [2] Deepayan Sarkar and Felix Andrews. *latticeExtra: Extra Graphical Utilities Based on Lattice*, 2016. URL <https://CRAN.R-project.org/package=latticeExtra>. R package version 0.6-28.
- [3] Rolf Turner. *deldir: Delaunay Triangulation and Dirichlet (Voronoi) Tessellation*, 2016. URL <https://CRAN.R-project.org/package=deldir>. R package version 0.1-12.
- [4] Fortran code by R. J. Renka. R functions by Albrecht Gebhardt. With contributions from Stephen Eglen <stephen@anc.ed.ac.uk>, Sergei Zuyev, and Denis White. *tripack: Triangulation of Irregularly Spaced Data*, 2015. URL <https://CRAN.R-project.org/package=tripack>. R package version 1.3-7.
- [5] Daniel Adler, Duncan Murdoch, and others. *rgl: 3D Visualization Using OpenGL*, 2016. URL <https://CRAN.R-project.org/package=rgl>. R package version 0.96.0.
- [6] Hadley Wickham. *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York, 2009. ISBN 978-0-387-98140-6. URL <http://ggplot2.org>.

Session Info

```
[,1]
sysname      "Linux"
release       "4.4.0-21-generic"
version       "#37-Ubuntu SMP Mon Apr 18 18:33:37 UTC 2016"
nodename     "cb-T61p"
machine      "x86_64"
login        "unknown"
user         "cb"
effective_user "cb"
```

```

R version 3.3.2 (2016-10-31)
Platform: x86_64-pc-linux-gnu (64-bit)
Running under: Ubuntu 16.04.1 LTS

locale:
[1] LC_CTYPE=de_DE.UTF-8          LC_NUMERIC=C           LC_TIME=de_DE.UTF-8
[4] LC_COLLATE=C                LC_MONETARY=de_DE.UTF-8   LC_MESSAGES=de_DE.UTF-8
[7] LC_PAPER=de_DE.UTF-8         LC_NAME=C              LC_ADDRESS=C
[10] LC_TELEPHONE=C             LC_MEASUREMENT=de_DE.UTF-8 LC_IDENTIFICATION=C

attached base packages:
[1] tools      grid       stats      graphics   grDevices  utils      datasets   methods    base

other attached packages:
[1] latticeExtra_0.6-28     RColorBrewer_1.1-2      baseline_1.2-1        MASS_7.3-45
[5] hyperSpec_0.98-20161118 ggplot2_2.2.0          lattice_0.20-34

loaded via a namespace (and not attached):
[1] Rcpp_0.12.7      magrittr_1.5      knitr_1.13      munsell_0.4.3    svUnit_0.7-12
[6] xtable_1.8-2     colorspace_1.3-0  R6_2.1.2       stringr_1.1.0    plyr_1.8.4
[11] gtable_0.2.0    plotrix_3.6-2     deldir_0.1-12   htmltools_0.3.5  lazyeval_0.2.0
[16] assertthat_0.1   digest_0.6.10    tibble_1.2       shiny_0.13.2    reshape2_1.4.2
[21] htmlwidgets_0.6  mime_0.4       labeling_0.3    stringi_1.1.2   scales_0.4.1
[26] SparseM_1.72    jsonlite_0.9.21  mvtnorm_1.0-5   httpuv_1.3.3

```